

# EasyGo!™ Human IL-10 One-Step ELISA Kit

[Catalog No] EK110EGB

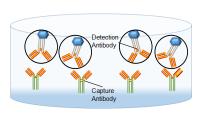
**[SIZE]** 48T/96T

[INTENDED USE] For the quantitative determination of human Interleukin10 (IL-10) concentrations in cell culture supernates.

#### [INTRODUCTION]

The IL-10 protein is a homodimer, each of its subunits is 178-amino-acid long. In humans, IL-10 is encoded by the IL10 gene, which is located on chromosome 1 and comprises 5 exons, and is primarily produced by monocytes and, to a lesser extent, lymphocytes, namely type 2 T helper cells (Th2), mastocytes, CD4+CD25+Foxp3+ regulatory T cells, and in a certain subset of activated T cells and B cells. IL-10 is a cytokine with multiple, pleiotropic, effects in immunoregulation and inflammation. It downregulates the expression of Th1 cytokines, MHC class II antigens, and co-stimulatory molecules on macrophages. It also enhances B cell survival, proliferation, and antibody production. IL-10 can block NF-KB activity, and is involved in regulation of the JAK-STAT signaling pathway. The immunosuppressive properties of IL-10 suggest a possible clinical use of IL-10 in suppressing rejections of grafts after organ transplantations. IL-10 can furthermore strona anti-inflammatory activities.

#### [PRINCIPLE OF THE ASSAY]







ELISA Plate Well Surface

Human IL-10 ELISA Kit is based on the quantitative sandwich enzyme-linked immunosorbent assay technique to measure concentration of human IL-10 in the samples. A monoclonal antibody specific for human IL-10 has been immobilized onto microwells, and one pellet of the HRP-linked detect antibody specific for IL-10 (light yellow) is pre-placed in the microwells, sealed by the adhesive film. Standard or samples are pipetted into the wells, then IL-10 present is bound by the immobilized antibody and detect antibody. After washing, substrate solution reacts with HRP and color develops in proportion to the amount of IL-10 bound by the immobilized antibody. The color development is stopped by addition of acid and the optical density value is measured by microplate reader.

# [MATERIALS PROVIDED]

PART	PART#	EK110EGB-48	EK110EGB-96
Coated Microplate	EK110EGBP	48T	96T
Standard	EK110EGBS	1 vial	2 vials
TMB	E0230	6 mL	11 mL
Stop Solution	E0300	11 mL	11 mL
Washing Buffer (20x)	F0281	11 ml	11 ml

Note: Components from reagent kits of different batch numbers must not be used interchangeably.

### OTHER SUPPLIES REQUIRED

- 1) Microplate reader capable of measuring absorbance at 450 nm, with correction wavelength set at 570 nm or 630 nm.
- 2) Pipettes and pipette tips.
- 3) 50  $\mu$ l to 300  $\mu$ l adjustable multichannel micropipette with

disposable tips.

- 4) Multichannel micropipette reservoir.
- 5) Beakers, flasks, cylinders necessary for preparation of reagents.
- 6) Deionized or distilled water.
- 7) Polypropylene test tubes for dilution.

#### [STORAGE]

Store at 2-8°C; refer to the kit label for expiration date.

For opened kits:

Pre-coated microplate: Can be stored at 2-8°C for approximately 1 month (return unused strips to the aluminum foil bag and reseal).

Standard: Can be stored at -20°C for approximately 1 month (discard after single-use reconstitution).

Other components: Can be stored at 2-8°C for approximately 1 month.

#### [SAMPLE COLLECTION AND STORAGE]

- 1) **Cell Culture Supernates** Remove particulates by centrifugation and assay freshly prepared samples immediately or aliquot and store samples at ≤ -20°C for later use. Avoid repeated freeze-thaw cycles.
- 2) Other biological samples might be suitable for use in the assay. Cell culture supernates were tested with this assay. Dilution with Assay Buffer may be needed.

Note: Samples containing a visible precipitate must be clarified prior to use in the assay. Do not use grossly hemolyzed or lipemic specimens.

If samples are to be run within 24 hours, they may be stored at 2 to 8°C. For longer storage, aliquot samples and store frozen at -20°C. Avoid repeated freeze-thaw cycles.

### [REAGENT PREPARATION]

Bring all reagents and samples to room temperature before use. If crystals form in the Buffer Concentrates, warm and gently stir them until completely dissolved.

## Washing Buffer (1×)

Pour entire contents (50 ml) of the **Washing Buffer (20x)** into a clean 1,000 ml graduated cylinder. Bring to final volume of 1,000 ml with pure or deionized water.

Mix gently to avoid foaming.

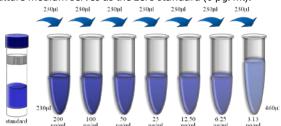
Transfer to a clean wash bottle and store at 2 to 25°C. Washing Buffer  $(1\times)$  is stable for 30 days.

**Sample Dilution:** If your samples have high **IL-10** content, dilute cell culture supernates with cell culture medium.

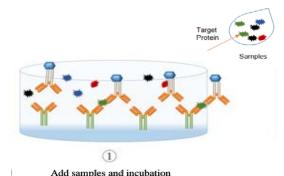
**Human IL-10 Standard**: Reconstitute **Human IL-10 Standard** by addition of distilled water. Reconstitution volume is stated on the label of the standard vial. Swirl or mix gently to insure complete and homogeneous solubilization (concentration of reconstituted standard = 400 pg/ml).

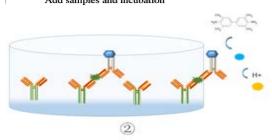
Allow the standard to reconstitute for 10 - 30 minutes. Mix well prior to making dilutions. Use polypropylene tubes.

• For cell culture supernates, mixing concentrated human IL-10 standard (230 µl) with 230 µl of cell culture medium creates the high standard (200 pg/ml). Pipette 230 µl of cell culture medium into each tube. Use the high standard to produce a 1:1 dilution series. Mix each tube thoroughly before the next transfer. Cell culture medium serves as the zero standard (0 pg/ml).



#### [ASSAY PROCEDURE]





Color developing and reading

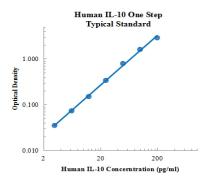
Bring all reagents and samples to room temperature before use.

- 1) Prepare all reagents including microplate, samples, standards and working solution as described in the previous sections.
- 2) Remove excess microplate strips with the knife and return them to the foil pouch containing the desiccant pack, and reseal for further use. In any case, avoid touching the inner surface of the microwells and gently tap the plate to ensure that the pellets on the bottom of the microwells. Do not discard the pellets.
- 3) Add 100  $\mu$ l 2-fold diluted Standard to Standard well. Add 100  $\mu$ l culture medium to Blank well. The standards/samples can be added directly from the middle opening of the plate adhesive film.
- 4) Cell culture supernates: Add 100  $\,\mu l$  cell culture supernates to the sample well.
- 5) Incubate at 37°C for **1** hour, or at room temperature (25°C±3°C) for **2** hours, on a microplate shaker set at 300-500 rpm.
- 6) Removing the plate adhesive film. Aspirate each well and wash by filling each well with 300  $\mu$ l Washing Buffer (1×), repeat three times for a total four washes with 60 seconds interval. Complete removal of liquid at each step is essential to the best performance. After the last wash, remove any remaining Washing Buffer (1×) by aspirating or decanting. Invert the plate and tap it against clean paper towels.
- 7) Add 100 µl of Substrate Solution to each well. Incubate for 10±5 minutes at room temperature(25°C±3°C). **Protect from light.**
- 8) Add 100  $\,\mu$ l of Stop Solution to each well. The color will turn yellow. If the color in the well is green or if the color change does not appear uniform, gently tap the plate to ensure thorough mixing.
- 9) Measure the optical density value within 30 minutes by microplate reader set to 450 nm. If wavelength correction is available, set to 570 nm or 630 nm. If wavelength correction is not available, subtract readings at 570 nm or 630 nm from the readings at 450 nm. This subtraction will correct for optical imperfections in the plate. Reading directly at 450 nm without correction may generate higher concentration than true value.

# [TYPICAL DATA]

A standard curve must be run within each assay. The following standard curve is provided for demonstration only.

	_	_	_	
pg/mL	0.D.		Average	Corrected
0.00	0.019	0.019	0.019	
3.13	0.056	0.052	0.054	0.035
6.25	0.091	0.093	0.092	0.073
12.50	0.172	0.165	0.169	0.150
25.00	0.366	0.347	0.357	0.338
50.00	0.786	0.830	0.808	0.789
100.00	1.574	1.671	1.623	1.604
200.00	3.023	2.766	2.895	2.876



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