

# EasyGo!™ Mouse IL-4 One-Step ELISA Kit

[Catalog No] EK204EGA

**(SIZE)** 48T/96T

[ INTENDED USE ] For the quantitative determination of mouse Interleukin 4 (IL-4) concentrations in serum and plasma.

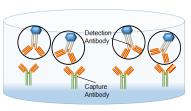
#### [INTRODUCTION]

Interleukin 4 (IL-4) is an approximately 18 - 20 kDa monomeric cytokine. Mouse IL-4 is synthesized as a 140 amino acid (aa) precursor with a 20 aa signal sequence and a 120 aa mature chain and mature mouse IL-4 shares 43 %, and 63 % aa sequence identity with human and rat IL-4.

IL-4 is a cytokine that induces differentiation of naive helper T cells (Th0 cells) to Th2 cells. Upon activation by IL-4, Th2 cells subsequently produce additional IL-4 in a positive feedback loop. The cell that initially produces IL-4, thus inducing Th0 differentiation, has not been identified, but recent studies suggest that basophils may be the effector cell. It is closely related and has functions similar to IL-13.

IL-4 also has been shown to drive mitogenesis, dedifferentiation and metastasis in rhabdomyosarcoma. IL-4, along with other Th2 cytokines, is involved in the airway inflammation observed in the lungs of patients with allergic asthma.

## [PRINCIPLE OF THE ASSAY]







ELISA Plate Well Surface

This assay employs the quantitative sandwich enzyme immunoassay technique. A monoclonal antibody specific for Mouse IL-4 has been immobilized onto microwells, and two pellets of the biotin-linked detect antibody specific for IL-4 (light yellow) and streptavidin-HRP (purple) are pre-placed in the microwells, sealed by the adhesive film. Standard or samples are pipetted into the wells, then IL-4 present is bound by the immobilized antibody and detect antibody, of which the latter is conjugated with streptavidin-HRP in the incubation. After washing, substrate solution reacts with HRP and color develops in proportion to the amount of IL-4 bound by the immobilized antibody. The color development is stopped and the intensity of the color is measured by microplate reader.

## [MATERIALS PROVIDED]

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PART	PART#	EK204EGA-	EK204EGA-			
FANI		48	96			
Coated Microplate	EK204EGAP	48T	96T			
standard	EK204EGAS	1 vial	2 vials			
Standard Diluent	E0260	5 mL	10 mL			
TMB	E0230	6 mL	11 mL			
Stop Solution	E0300	11 mL	11 mL			
Washing Buffer (20×)	F0281	11 ml	11 ml			

Note: Components from reagent kits of different batch numbers must not be used interchangeably.

# OTHER SUPPLIES REQUIRED

1) Microplate reader capable of measuring absorbance at 450 nm,

with correction wavelength set at 570 nm or 630 nm.

- 2) Pipettes and pipette tips.
- 3) 50  $\,\mu$  l to 300  $\,\mu$  l adjustable multichannel micropipette with disposable tips.
- 4) Multichannel micropipette reservoir.
- 5) Beakers, flasks, cylinders necessary for preparation of reagents.
- 6) Deionized or distilled water.
- 7) Polypropylene test tubes for dilution.

#### [STORAGE]

Store at 2-8°C; refer to the kit label for expiration date.

For opened kits:

Pre-coated microplate: Can be stored at 2-8°C for approximately 1 month (return unused strips to the aluminum foil bag and reseal). Standard: Can be stored at -20°C for approximately 1 month (discard after single-use reconstitution).

Other components: Can be stored at 2-8°C for approximately 1 month.

#### [SAMPLE COLLECTION AND STORAGE]

- 1) **Serum** Use a serum separator tube (SST) and allow samples to clot for 30 minutes before centrifugation for 10 minutes at 1,000  $\times$  g. Remove serum and assay freshly prepared samples immediately or aliquot and store samples at  $\leq$  -20°C for later use. Avoid repeated freeze-thaw cycles.
- 2) Plasma Collect plasma using EDTA, citrate or heparin as anticoagulant. Centrifuge at 1,000 × g within 30 minutes of collection. Assay freshly prepared samples immediately or aliquot and store samples at ≤ -20°C for later use. Avoid repeated freeze-thaw cycles.
   3) Other biological samples might be suitable for use in the assay.
- Serum and plasma were tested with this assay. Dilution with Standard Diluent may be needed.

  Note: Samples containing a visible precipitate must be clarified prior

to use in the assay. Do not use grossly hemolyzed or lipemic specimens.

If samples are to be run within 24 hours, they may be stored at 2 to 8°C. For longer storage, aliquot samples and store frozen at -20°C. Avoid repeated freeze-thaw cycles.

# [REAGENT PREPARATION]

Bring all reagents and samples to room temperature before use. If crystals form in the Buffer Concentrates, warm and gently stir them until completely dissolved.

### Washing Buffer (1×)

Pour entire contents (50 ml) of the **Washing Buffer (20x)** into a clean 1,000 ml graduated cylinder. Bring to final volume of 1,000 ml with pure or deionized water.

Mix gently to avoid foaming.

Transfer to a clean wash bottle and store at 2 to 25°C. Washing Buffer (1x) is stable for 30 days.

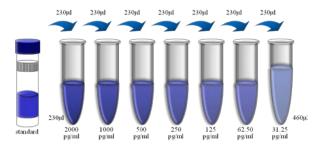
**Sample Dilution:** If your samples have high **IL-4** content, dilute serum/plasma samples with Standard Diluent.

Mouse IL-4 Standard: Reconstitute Mouse IL-4 Standard by addition of distilled water. Reconstitution volume is stated on the label of the standard vial. Swirl or mix gently to insure complete and homogeneous solubilization (concentration of reconstituted standard = 4,000 pg/ml).

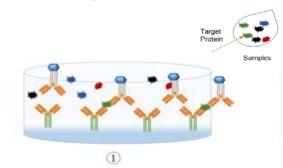
Allow the standard to reconstitute for 10 - 30 minutes. Mix well prior to making dilutions. Use polypropylene tubes.

• For serum/plasma samples, mixing concentrated Mouse IL-4 standard (230 μl) with 230 μl of Standard Diluent creates the high standard (2,000 pg/ml). Pipette 230 μl of Standard Diluent into each tube. Use the high standard to produce a 1:1 dilution series

(scheme below). Mix each tube thoroughly before the next transfer. Standard Diluent serves as the zero standard (0 pg/ml).



## [ASSAY PROCEDURE]



Add samples and incubation

Color developing and reading

Bring all reagents and samples to room temperature before use.

- 1) Prepare all reagents including microplate, samples, standards and working solution as described in the previous sections.
- 2) Remove excess microplate strips and return them to the foil pouch containing the desiccant pack, and reseal for further use. In any case, avoid touching the inner surface of the microwells and gently tap the plate to ensure that the pellets on the bottom of the microwells. Do not discard the pellets.
- 3) Adding Standard: Add 100  $\mu$ L of 2-fold diluted Standard to Standard well. Add 100  $\mu$ L of Standard Diluent to Blank well. The standards/samples can be added directly from the middle opening of the plate adhesive film.
- 4) Adding Samples: Add 90  $\,\mu L$  of Standard Diluent and 10  $\,\mu L$  sample to the sample well.
- 5) Incubation: Incubate at 37°C for **1** hour, or at room temperature (25°C  $\pm$ 3°C) for **2** hours, on a microplate shaker set at 300-500 rpm.
- 6) Washing: Removing the plate adhesive film. Aspirate each well and wash by filling each well with 300µL of Washing Buffer (1×), repeating the process 3 times for a total four washes with 60 seconds interval. Complete removal of liquid at each step is essential to good performance. After the last wash, remove any remaining Wash Buffer by aspirating or decanting. Invert the plate and blot it against clean paper towels.
- 7) Adding Substrate Solution: Add 100 µL of Substrate Solution to each well. Incubate for 10±5 minutes at room temperature (25±3°C). **Protect from light.**
- 8) Stopping: Add 100  $\mu$ L of Stop Solution to each well. The color will turn yellow. If the color in the well is green or if the color change does

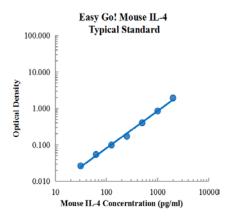
not appear uniform, gently tap the plate to ensure thorough mixing.

9) Reading: Measure the optical density value within 30 minutes by microplate reader set to 450 nm. If wavelength correction is available, set to 570 nm or 630 nm. If wavelength correction is not available, subtract readings at 570 nm or 630 nm from the readings at 450 nm. This subtraction will correct for optical imperfections in the plate. Reading directly at 450 nm without correction may generate higher concentration than true value.

## [TYPICAL DATA]

A standard curve must be run within each assay. The following standard curve is provided for demonstration only.

pg/mL	O.D.		Average	Corrected	
0.00	0.062	0.062	0.062		
31.25	0.097	0.080	0.089	0.027	
62.50	0.116	0.117	0.117	0.055	
125.00	0.161	0.160	0.161	0.099	
250.00	0.238	0.229	0.234	0.172	
500.00	0.447	0.482	0.465	0.403	
1000.00	0.985	0.836	0.911	0.849	
2000.00	1.959	2.029	1.994	1.932	



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