

Mouse Total HIF-1α/HIF1A ELISA Kit

[Catalog No] EK2392

[SIZE] 48T/96T

[INTENDED USE] For the quantitative detection of mouse hypoxiainducible transcription factor 1α (HIF- 1α /HIF1A) concentration in cell lysates.

[INTRODUCTION]

Hypoxia-inducible factor 1α (HIF- 1α /HIF1A) is the regulatory subunit of the transcription factor heterodimer HIF-1 $\alpha\,/\,\beta$. As the major transcriptional regulator of adaptive responses to hypoxia, under hypoxic conditions, HIF-1 α binds to hypoxia-response elements (HREs) in the promoters of many genes, activating the transcription of more than 40 genes including erythropoietin, glucose transporters, glycolytic enzymes, vascular endothelial growth factor, HILPDA, and other genes whose protein products increase oxygen delivery or promote metabolic adaptation to hypoxia. HIF-1 α also plays an important role in the pathophysiology of embryonic angiogenesis, tumor angiogenesis, and ischemic diseases, as well as participates in the axonal distribution and transport of mitochondria within neurons during hypoxia. HIF-1 α is ubiquitously present in human and mammalian cells, and it is expressed even under normoxic conditions (21% O_2), but the synthesized HIF-1 α protein is rapidly degraded by the intracellular oxygen-dependent ubiquitin-proteasome pathway. Only under hypoxic conditions can HIF-1 α be stably expressed.

[PRINCIPLE OF THE ASSAY]

[MATERIALS PROVIDED]

This assay employs the quantitative sandwich enzyme immunoassay technique. A monoclonal antibody specific for mouse Total HIF-1 α /HIF1A has been pre-coated onto a microplate. Standard, samples and biotin-linked detect antibody specific for Total HIF-1 α /HIF1A are pipetted into the wells and Total HIF-1 α /HIF1A present is bound by the immobilized antibody and detect antibody following incubation. After washing away any unbound substances, streptavidin-HRP is added. After washing, substrate solution is added to the wells and color develops in proportion to the amount of Total HIF-1 α /HIF1A bound in the initial step. The color development is stopped and the intensity of the color is measured.

PART	PART #	EK2392-48	EK2392-96		
Coated Microplate	EK2392P	48T	96T		
Standard	EK2392S	1 vial	2 vials		
Detect antibody	EK2392D	1 vial	1 vial		
Standard Diluent	E0260	5 ml	5 ml		
Streptavidin-HRP	E0290	1 vial	1 vial		
Assay Buffer (10×)	E0310	5ml	5ml		
ТМВ	E0230	6ml	11ml		
Stop Solution	E0300	11ml	11ml		
Washing Buffer (20×)	E0281	50ml	50ml		
Adhesive Film	E0200	6	6		

Note: Components from reagent kits of different batch numbers must not be used interchangeably.

OTHER SUPPLIES REQUIRED

1) Microplate reader capable of measuring absorbance at 450 nm, with correction wavelength set at 570 nm or 630 nm.

2) Pipettes and pipette tips.

3) 50 $\,\mu\,l\,$ to 300 $\,\mu\,l\,$ adjustable multichannel micropipette with

disposable tips.

4) Multichannel micropipette reservoir.

5) Beakers, flasks, cylinders necessary for preparation of reagents.

6) Deionized or distilled water.

7) Polypropylene test tubes for dilution.

[STORAGE]

Store at 2-8°C; refer to the kit label for expiration date. For opened kits:

Pre-coated microplate: Can be stored at 2-8°C for approximately 1 month (return unused strips to the aluminum foil bag and reseal). Standard: Can be stored at -20°C for approximately 1 month (discard

after single-use reconstitution).

Other components: Can be stored at 2-8°C for approximately 1 month. [SAMPLE COLLECTION AND STORAGE]

1) **Cell Lysate:** Rinse cells twice with PBS, ensuring that any remaining PBS is removed after the second rinse. Add cell lysis buffer

(self-prepared) at a concentration of 1×10^7 cells/ml. Place on ice for 15 minutes and then collect samples for immediate measurement or store at \leq -20°C. Prior to use, centrifuge the sample at 2,000×g for 5 minutes and transfer the supernatant to a clean tube.

2) Other biological samples might be suitable for use in the assay. Cell Lysates were tested with this assay.

Note: Samples containing a visible precipitate must be clarified prior to use in the assay. Do not use grossly hemolyzed or lipemic specimens.

If samples are to be run within 24 hours, they may be stored at 2 to 8°C. For longer storage, aliquot samples and store frozen at -20°C. Avoid repeated freeze-thaw cycles.

[REAGENT PREPARATION]

Bring all reagents and samples to room temperature before use. If crystals form in the Buffer Concentrates, warm and gently stir them until completely dissolved.

Washing Buffer (1×)

Pour entire contents (50 ml) of the **Washing Buffer (20x)** into a clean 1,000 ml graduated cylinder. Bring to final volume of 1,000 ml with pure or deionized water.

Mix gently to avoid foaming.

Transfer to a clean wash bottle and store at 2 to 25° C. Washing Buffer (1×) is stable for 30 days.

Assay Buffer (1×)

Pour the entire contents (5 ml) of the **Assay Buffer (10x)** into a clean 100 ml graduated cylinder. Bring to final volume of 50 ml with distilled water. Mix gently to avoid foaming.

Store at 2 to 8°C. Assay Buffer (1×) is stable for 30 days.

Detect Antibody

Mix well prior to making dilutions.

Make a 1: 100 dilution of the concentrated **Detect Antibody** solution with Assay Buffer $(1\times)$ in a clean plastic tube.

The diluted Detect Antibody should be used within 30 minutes after dilution.

Streptavidin-HRP

Mix well prior to making dilutions.

Make a **1: 100** dilution of the concentrated **Streptavidin-HRP** solution with Assay Buffer $(1\times)$ in a clean plastic tube as needed.

The diluted Streptavidin-HRP should be used within 30 minutes after dilution.

Sample Dilution: If your samples have high Total HIF-1 α /HIF1A content, dilute samples with Assay Buffer (1×).

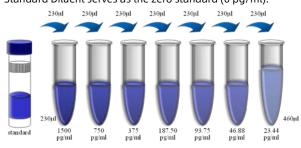
Mouse Total HIF-1 α **/HIF1A Standard:** Briefly spin the vial at 6,000 rpm

for 30 seconds before opening. Reconstitute mouse Total HIF-1 α /HIF1A Standard by addition of distilled water. Reconstitution volume is stated on the label of the standard vial. Swirl or mix gently to insure complete and homogeneous solubilization (concentration of reconstituted standard = 3,000 pg/ml).

Allow the standard to reconstitute for 10 - 30 minutes. Mix well prior to making dilutions.

Use polypropylene tubes.

For cell lysates, mixing concentrated mouse Total HIF-1α/HIF1A standard (230 μl) with 230μl of Standard Diluent creates the high standard (1,500 pg/ml). Pipette 230 μl of Standard Diluent into each tube. Use the high standard to produce a 1:1 dilution series (scheme below). Mix each tube thoroughly before the next transfer. Standard Diluent serves as the zero standard (0 pg/ml).



[ASSAY PROCEDURE]

Bring all reagents and samples to room temperature before use. 1) Prepare all reagents including microplate, samples, standards and working solution as described in the previous sections.

2) Remove excess microplate strips and return them to the foil pouch containing the desiccant pack, and reseal for further use.

3) Add 300 μ l Washing Buffer (1×) per well, and allow it for about 30 seconds before aspiration. Soaking is highly recommended to obtain a good test performance. Empty wells and tap microwell strips on absorbent pad or paper towel to remove excess Washing Buffer (1×). Use the microwell strips immediately after washing. **Do not allow wells to dry.**

4) Add 100 μl 2-fold diluted Standard to Standard well. Add 100 μl Standard Diluent/ culture medium to Blank well.

5) Add 90 $\,\mu l$ Assay Buffer (1×) and 10 $\,\mu l$ sample to the sample well.

6) Add 50 μl of diluted Detect Antibody to each well. Ensure reagent addition in step 4, 5 and 6 is uninterrupted and completed within 15 minutes.

7) **Seal the plate with an adhesive film.** Incubate at room temperature (25°C±3°C) for 2 hours on a microplate shaker set at 300 rpm.

8) Aspirate each well and wash by filling each well with 300 μ l Washing Buffer (1×), repeat five times for a total six washes. Complete removal of liquid at each step is essential to the best performance. After the last wash, remove any remaining Washing Buffer (1×) by aspirating or decanting. Invert the plate and tap it against clean paper towels.

9) Add 100 µl of diluted Streptavidin-HRP to each well.

10) **Seal the plate with a fresh adhesive film.** Incubate at room temperature (25°C±3°C) for 45 minutes on a microplate shaker set at 300 rpm.

11) Repeat aspiration/wash as in step 8.

12) Add 100 µl of Substrate Solution to each well. Incubate for 5 - 30 minutes at room temperature(25°C±3°C). **Protect from light.**

13) Add 100 µl of Stop Solution to each well. The color will turn yellow. If the color in the well is green or if the color change does not appear uniform, gently tap the plate to ensure thorough mixing.

14) Measure the optical density value within 30 minutes by microplate

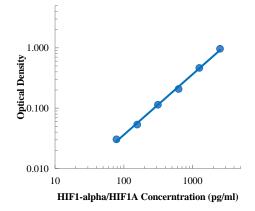
reader set to 450 nm. If wavelength correction is available, set to 570 nm or 630 nm. If wavelength correction is not available, subtract readings at 570 nm or 630 nm from the readings at 450 nm. This subtraction will correct for optical imperfections in the plate. Reading directly at 450 nm without correction may generate higher concentration than true value.

[TYPICAL DATA]

A standard curve must be run within each assay. The following standard curve is provided for demonstration only.

pg/ml	0.D.		Average	Corrected
0.00	0.016	0.074	0.045	
23.44	0.064	0.063	0.064	0.019
46.88	0.108	0.106	0.107	0.062
93.75	0.195	0.205	0.200	0.155
187.50	0.338	0.378	0.358	0.313
375.00	0.645	0.658	0.652	0.607
750.00	1.194	1.195	1.195	1.150
1,500.00	2.374	2.393	2.384	2.339





MULTI SCIENCES BIOTECH CO., LTD. www.multisciences.net E-mail: service@multisciences.net